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(19) (CA) **APPLICATION FOR CANADIAN PATENT** (12)

(54) Glycerin-3-Phosphate-Dehydrogenase (GPDH)

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(30) (DE) P 43 29 827.3 1993/09/03

(57) 12 Claims

Notice: This application is as filed and may therefore contain an
incomplete specification.



MAX PLANCK SOCIETY

for Promotion of the Sciences e.V. [Registered Association]

37073 Goettingen

Glycerol-3-phosphate dehydrogenase (GPDH)

This invention concerns DNA sequences that code for a glycerol-3-phosphate dehydrogenase (GPDH) and the alleles as well as the derivatives of these DNA sequences.

This invention also concerns genomic clones that contain the complete gene of a glycerol-3-phosphate dehydrogenase and alleles as well as derivatives of this gene.

This invention also concerns promoters and other regulator elements of glycerol-3-phosphate dehydrogenase genes.

Glycerol-3-phosphate dehydrogenase (GPDH; EC 1.1.1.8), also known as dihydroxyacetone phosphate reductase, is substantially involved in triacylglyceride biosynthesis in plants by supplying glycerol-3-phosphate. Fatty acid biosynthesis and triacylglyceride biosynthesis can be regarded as separate biosynthesis pathways owing to compartmentalization but as one biosynthesis pathway from the standpoint of the end product. *De novo* biosynthesis of fatty acids takes place in the plastids and is catalyzed by three enzymes or enzyme systems, i.e., (1) acetyl-CoA carboxylase (ACCase), (2) fatty acid synthase (FAS), and (3) acyl-[ACP]-thioesterase (TE). The end products of this reaction sequence in most organisms are either palmitic acid, stearic acid, or after desaturation, oleic acid.

In the cytoplasm, however, triacylglyceride biosynthesis takes place via the so-called "Kennedy pathway" in the endoplasmic reticulum from glycerol-3-phosphate which is made available by the activity of glycerol-3-phosphate dehydrogenase (S.A. Finnlayson et al., Arch. Biochem. Biophys., 199 (1980)

pages 179-185), and from fatty acids present in the form of acyl-CoA substrates.

Probably the first discovery of the enzymatic activity of glycerol-3-phosphate dehydrogenase in plants involved potato tubers (G.T. Santora et al., Arch. Biochem. Biophys., 196 (1979) pages 403-411). This activity had not been observed in other plants before then (B. König and E. Heinz, Planta, 118 (1974) pages 159-169), so the existence of the enzyme had not been detected. Thus the formation of glycerol-3-phosphate on the basis of the activity of a glycerol kinase was discussed as an alternative biosynthesis pathway. Santora et al., loc. cit., subsequently detected GPDH in spinach leaves and succeeded in increasing the concentration of the enzyme approximately 10,000 times. They determined the native molecular weight to be 63.5 kDa and found the optimum pH for the reduction of dihydroxyacetone phosphate (DHAP) to be 6.8 to 9.5 for the back reaction. GPDH was likewise detected in Ricinus endosperm (Finlayson et al., Biochem. Biophys. 199 (1980) pages 179-185). According to more recent works (Gee et al., Plant Physiol. 86 (1988a) pages 98-103), two GPDH activities could be detected in enriched fractions, a cytoplasmic fraction (20-25%) and a plastid (75-80%). The two forms are regulated differently. Thus, for example, the cytoplasmic isoform can be activated by F₂,6DP, while the plastid isoform is activated by thioredoxin (R.W. Gee et al., Plant Physiol., 86 (1988) pages 98-103 and R.W. Gee et al., Plant Physiol., 87 (1988) pages 379-383).

The methods of molecular biology are making increasing entry into plant cultivation practice. Changes in biosynthesis output with the formation of new components and/or higher yields of these components can be achieved with the help of gene manipulation, e.g., transfer of genes which code for enzymes. As one of the most important enzymes of triacylglyceride synthesis, GPDH has a significant influence on the oil yield of plants.

It is thus the object of this invention to improve the oil yield of crop plants by influencing the triacylglyceride content.

This object is achieved with the DNA sequences according to patent claim 1 and the genes from the genomic clones according to patent claim 4.

This invention concerns DNA sequences that code for a glycerol-3-phosphate dehydrogenase, and alleles as well as derivatives of these DNA sequences.

This invention also concerns genomic clones that contain a complete gene of a glycerol-3-phosphate dehydrogenase including the structure gene, the promoter and other regulator sequences, and alleles as well as derivatives of this gene.

This invention likewise concerns the promoters and other regulator elements of glycerol-3-phosphate dehydrogenase genes from the specified genomic clones, and the alleles as well as derivatives of these promoters.

This invention additionally concerns a method of producing plants, plant parts and plant products in which the triacylglyceride content or fatty acid content is altered, where DNA sequences or genes are transferred from the genomic clones by the methods of genetic engineering.

This invention also concerns the use of said DNA sequences or one of the genes originating from said genomic clones for altering the triacylglyceride content or its fatty acid pattern in plants.

Finally, this invention concerns transgeneic plants, plant parts and plant products produced according to the aforementioned method.

The figures serve to clarify the present invention.

They show the following:

- Figure 1: Comparison of the derived amino acid sequences of the ClGPDH30 and ClGPDH109 cDNAs as well as the gene from the ClGPDHg3 genomic clone with the GPDH amino acid sequence of the mouse (Mm GPDH);
- Figure 2: Separation of proteins from BB26-36 cells by gel electrophoresis;
- Figure 3: Map of the insertions contained in ClGPDHg5, ClGPDHg9 and ClGPDHg3 genomic clones with various restriction enzymes;
- Figure 4: Schematic diagram of the functional areas of the genes contained in the ClGPDH5, ClGPDH9 and ClGPDH3 genomic clones; and
- Figure 5: Northern Blot with RNAs from various plant tissues, hybridized with ClGPDH20 cDNA as a probe.

It is obvious that allelic variants and derivatives of DNA sequences or genes according to this invention are included within the scope of this invention under the assumption that these modified DNA sequences or modified genes will code for glycerol-3-phosphate dehydrogenase. The allelic variants and derivatives include, for example, deletions, substitutions, insertions, inversions and additions to DNA sequences or genes according to this invention.

Any plant material that produces glycerol-3-phosphate dehydrogenase in sufficient quantities is a suitable raw material for isolating cDNAs that code for glycerol-3-phosphate dehydrogenase. Isolated embryos from the plant *Cuphea lanceolata*, indigenous to Central America, have proven to be an especially suitable raw material in the present invention.

Functional complementation was used for isolation of DNA sequences according to this invention. This refers to complementation of mutant microorganisms with heterologous cDNA. Functional complementation was performed after infecting *E. coli* strain BB26-36, which is auxotrophic for glycerol, with phagemids containing plasmids with cDNAs from *Cuphea lanceolata*.

Plasmids isolated from functionally complemented bacteria were cleaved with

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restriction endonucleases and separated by electrophoresis. The cDNAs contained in the plasmids were classified in two classes that differ in the size of their insertions. Retransformation confirmed that the isolated cDNAs were capable of complementing the BB26-36 mutant.

The complete coding area of one of the two classes codes for a glycerol-3-phosphate dehydrogenase and is contained in the ClGPDH20 cDNA clone. This is an Eco RI-ApaI fragment that has 1354 base pairs. The complete 1354 base pair DNA sequence of the ClGPDH20 cDNA and the amino acid sequence derived from it are entered in the Sequence Listing as SEQ ID NO:1. ClGPDH20 cDNA was sequenced double stranded. Proceeding from the ATG start codon, the cDNA codes from positions 17 to 1132 for a protein with 372 amino acids (ending at the TAG stop codon), which is expressed as a fusion with lacZ without a shift in the reading frame. The estimated molecular weight is 40.8 kDa. Two base pairs (CA) preceding ATG are included with the cDNA. The first 14 nucleotides are attributed to the DNA sequence of the fusion with lacZ, and the linker sequence is indicated at the 3' end. The polyA signal is found at positions 1329 to 1334 in the 3' untranslated region.

It is assumed that ClGPDH20 cDNA is a cytoplasmic isoform, because no transit peptide can be detected in homology comparisons with mouse GPDH (see Figure 1). On the basis of the position of an assumed NADH binding site corresponding to the consensus sequence GxGxxG (see positions 29 to 34 in the ClGPDH20 amino acid sequence in Figure 1 (R.K. Wierenga et al., Biochem. 24 (1985) pages 1346-1357), the N-terminal sequence of 28 amino acids is not sufficient to code for a transit peptide whose length varies between 32 and 75 amino acids (Y. Gavel et al., FEBS Lett. 261 (1990) pages 455-458).

A cDNA library from *Cuphea lanceolata* was screened with ClGPDH20 cDNA as a probe for isolation of additional GPDH cDNAs, and a total of 52 cDNA clones

were isolated. - The 18 longest cDNAs were completely or partially sequenced. The ClGPDH109, ClGPDH30 and ClGPDH132 cDNA clones contain cDNAs with the complete coding region or a virtually complete cDNA of GPDH.

The ClGPDH109 cDNA clone contains the complete coding region of GPDH on a 1464 base pair EcoRI-ApaI DNA fragment which codes for a protein with 381 amino acids. The DNA sequence and the amino acid sequence derived from it are shown as SEQ ID NO:2 in the Sequence Listing. The DNA fragment was sequenced double stranded. The coding area begins with the ATG start codon in position 45 and ends in position 1187, followed by the TAG stop codon (positions 1188 to 1190). The cDNA itself begins at position 15. The first 14 nucleotides are attributed to the DNA sequence of the fusion with lacZ. The polyA signal (positions 1414 to 1419) and the polyA area (positions 1446 to 1454) as well as the linker sequence (positions 1459 to 1464) are found in the untranslated region at the 3' end.

Another cDNA, ClGPDH30, also contains the complete coding region of GPDH on a 1390 base pair EcoRI-XhoI fragment, which codes for a protein with 372 amino acids. The double-stranded-sequenced DNA sequence and the DNA sequence derived from it are listed as SEQ ID NO:4 in the Sequence Listing. The protein coding sequence begins with the ATG start codon at position 34 and ends before the stop codon at position 1149. The first 14 base pairs are attributed to the sequence of the fusion with lacZ. The polyA signal (positions 1349 to 1354) and the polyA region (positions 1366 to 1384) are found in the untranslated 3' area.

The ClGPDH132 cDNA clone with 1490 base pairs is an Eco RI-XhoI fragment, the DNA sequence of which and the amino acid sequence derived from it are shown as SEQ ID NO:3 in the Sequence Listing. The DNA fragment was sequenced double stranded. ClGPDH132 cDNA is missing 14 amino acids at the N terminus In

comparison with ClGPDH109 cDNA. The open reading frame begins at position 15 and ends at position 1115, followed by the stop codon at positions 1116 to 1118. Consequently, ClGPDH132 cDNA codes for a protein with 367 amino acids and likewise includes the coding area for glycerol-3-phosphate dehydrogenase with the exception of 14 amino acids. The first 14 nucleotides are to be attributed to the lacZ fusion sequence and the linker sequence (positions 1485 to 1490) is at the 3' end. The polyA signal and the polyA area are located at positions 1343 to 1348 and 1465 to 1484, respectively, in the untranslated 3' area.

Two classes of cDNAs can be distinguished on the basis of sequence data. Accordingly, ClGPDH20 and ClGPDH30 cDNAs belong to class A and ClGPDH132 and ClGPDH109 cDNAs belong to class B.

As Figure 1 shows, the derived amino acid sequences of ClGPDH30 and ClGPDH109 cDNAs show 96% identical amino acids. At the same time, the derivative amino acid sequences of the cDNAs and those of a gene to be assigned to another class, ClGPDH30, were compared with the GPDH amino acid sequence of the mouse (MmGPDH). The differences between the amino acid sequence derived from the ClGPDH109 cDNA, the coded amino acid sequence of the gene and the mouse GPDH in comparison with the amino acid sequence derived from ClGPDH30 are shown in black. On the average, the identity of the derivative proteins of the cDNAs and the GPDH gene with the mouse protein is approximately 50%.

ClGPDH20 cDNA was cloned into an expression vector and expressed in *E. coli* as a fusion protein with glutathione-S-transferase. To do so, the cDNA was cloned beginning with ATG (see position 17, SEQ ID NO:1) into pGX, a derivative of the pGEXKG expression vector (K.L. Guan et al., *Analytical Biochem.* 192 (1991) pages 262-267). BB26-36 cells were harvested at various times after administration of IPTG (isopropyl-b-thiogalactopyranoside) and

their proteins were separated by gel electrophoresis. Figure 2 shows gel electrophoretic separation of BB26-36 cell extracts. The left column shows the proteins of cells with the pGX expression vector (without fusion; 26 kDa protein) and the right side shows proteins of cells with the pGXGPDH20 expression vector which codes for a fusion protein of 67 kDa. The hourly values given indicate the times of sampling after IPTG induction. This clearly shows an enrichment of the fusion protein after two hours. An enzyme activity determination was subsequently performed by enzyme assay of GPDH with an isolated fusion protein and significant enzyme activity was measured. This finding clearly proves that ClGPDH20 cDNA contains a competent gene for expression of GPDH.

Furthermore, genomic clones were isolated, where a library of genomic DNA of *Cuphea lanceolata* was screened with ClCPDH20 cDNA as a probe. By this method, 31 genomic clones were isolated. The genomic clones contain a complete structure gene of a glycerol-3-phosphate dehydrogenase and alleles plus derivatives of this gene together with the promoter sequence and other regulator elements. This means that they form complete transcription units.

Three genomic clones are characterized below. These include the ClGPDHg3 genomic clone with a 15.9 kb DNA insertion, the ClGPDHg5 genomic clone with a 17.7 kb DNA insertion, and the ClGPDHg9 genomic clone with a 15.6 kb DNA insertion. Figure 3 shows a map of the DNA insertions of the genomic clones with various restriction enzymes. The black bars indicate the fragments that hybridize with a 5' probe of the GPDH20 cDNA. The white bars show the areas of DNA insertions that were sequenced and are included in the Sequence Listing.

Sequence analysis of the areas presented in Figure 3 (white bars) of the three genomic clones ClGPDHg5, ClGPDHg3 and ClGPDHg9 has shown that they

contain the complete or partial structure gene of GPDH with all or most of the promoter sequence (5' direction). Figure 4 shows a schematic diagram of the sequenced areas of the genomic clones. The ClGPDHg5, ClGPDHg9 and ClGPDHg3 genomic clones contain the complete structure genes of GPDH in addition to promoter sequences. The entire promoter of GPDH was sequenced from the ClGPDHg9 genomic clone.

Thus a 4434 bp DNA fragment of the ClGPDHg5 genomic clone contains parts of the promoter and the complete structure gene of GPDH in the 5' area. The double-stranded-sequenced DNA sequence as well as the amino acid sequence derived from it are shown as SEQ ID NO:5 in the Sequence Listing. The protein-coding sequence interrupted by DNA areas not translated (introns) with 372 amino acids begins with the ATG start codon in position 1394 and ends before the TAG stop codon in position 4005. The putative TATA box is located at positions 1332 to 1336. Transcription presumably starts at position 1364 (Joshi, NAR 15 (1987) pages 6643-6653). The polyA signal is located in positions 4205 to 4210 at the 3' end. Position 4221 corresponds to the last nucleotide before the polyA area of ClGPDH30 cDNA (see position 1365 in SEQ ID NO:4).

The complete structure gene of GPDH as well as parts of the promoter in 5' direction are contained in a 4006 bp DNA fragment from the ClGPDHg3 genomic clone. The DNA sequence of the DNA fragment that was sequenced mostly as a double strand from ClGPDHg3 as well as the amino acid sequence derived from it are shown as SEQ ID NO:6a and SEQ ID NO:6b in the Sequence Listing. The protein coding area interrupted by intron sequences begins at position 1182 (see SEQ ID NO:6a) with the ATG start codon and ends with the TAG stop codon at position 190 (see SEQ ID NO:6b). CAAT box and TATA box signal sequences are located at positions 1055 to 1058 and 1103-1107 before the start of

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transcription. Assumed transcription starting points are at positions 1136 and 1148. Owing to a lack of sequence data, an area of approximately 480 base pairs is not identified within the coding sequence. The polyA signal is located in the untranslated 3' area at positions 393 to 398 (SEQ ID NO:6b).

The entire promoter as well as the first exon of the sequence coding for GPDH are contained in a 1507 bp DNA fragment from the ClGPDHg9 genomic clone. The DNA sequence that was sequenced mostly as a double strand as well as the amino acid sequence derived from it are shown as SEQ ID NO:7 in the Sequence Listing. The TATA box is located at positions 1108 to 1112 before the start of transcription. The protein coding sequence begins with the ATG start codon at position 1193 and ends at position 1376, where an untranslated area (intron) begins. Transcription presumably starts at position 1144.

By comparing DNA sequences, it has been found that ClGPDH30 cDNA, which includes a complete protein reading frame for GPDH, is identical to the GPDH gene from the ClGPDHg5 genomic clone. Consequently, the ClGPDHg5 genomic clone can be classified in class A (see above). The ClGPDH132 cDNA with an almost complete protein reading frame for GPDH is identical to the gene from the ClGPDHg9 genomic clone, which consequently may be assigned to class B (see above). The gene from the ClGPDHg3 genomic clone cannot be assigned to either of the two classes, and thus forms another class C.

Genetic engineering methods (in the form of anti-sense expression or overexpression) can be used to introduce or transfer the DNA sequences according to this invention that code for a glycerol-3-phosphate dehydrogenase into plants for the production of these dehydrogenases for the purpose of altering the biosynthesis yield of these plants. Inasmuch as the DNA sequences according to this invention are not a complete transcription unit, they are preferably introduced into the plants together with suitable promoters,

especially in recombinant vectors, such as binary vectors. Genomic clones can be used as separate complete transcription units for the transformation of plants in order to influence the triacylglyceride content and the fatty acid distribution.

Any species of plants can be transformed for this purpose. Oil-bearing plants, such as rapeseed, sunflower, linseed, oil palm and soybean are preferred for this transformation in order to influence the triacylglyceride biosynthesis in these plants in the manner desired.

The introduction of DNA sequences according to this invention that code for a glycerol-3-phosphate dehydrogenase as well as the complete genes contained in the genomic clones of a glycerol-3-phosphate dehydrogenase by the methods of genetic engineering can be performed with the aid of conventional transformation techniques. Such techniques include direct gene transfer, such as microinjection, electroporation, use of particle gun, steeping plant parts in DNA solutions, pollen or pollen tube transformation, viral vector-mediated transfer and liposome-mediated transfer as well as the transfer of appropriate recombinant Ti plasmids or Ri plasmids through *Agrobacterium tumefaciens* and transformation by plant viruses.

The DNA sequences according to this invention as well as the complete genes of a glycerol-3-phosphate dehydrogenase contained in the genomic clones are excellent for achieving a significant increase in oil production by transgeneic plants. This increase in oil yield is obtained with an increase in triacylglyceride content in of the seed due to overexpression of GPDH. Furthermore, a reduction in glycerol-3-phosphate dehydrogenase can be obtained through anti-sense expression or cosuppression, so the building blocks for triacylglyceride synthesis are missing. This effect is especially beneficial when the production of wax esters (such as jojoba wax esters) in the seeds of

transgeneic plants is to be improved. Another possible application of DNA sequences according to this invention as well as the genes from the genomic clones would be for suppressing triacylglyceride biosynthesis in transgeneic plants and making available the CoA ester as well as glycerol-3-phosphate for other biosyntheses.

Moreover, the promoters of glycerol-3-phosphate dehydrogenase genes from clones according to this invention can, for example, be used for targeted expression of chimeric genes in embryo-specific tissue. On the basis of experimental data it is assumed with regard to the specificity of the promoters that the promoters of genes from the ClGPDHg5 and ClGPDHg9 genomic clones are seed-specific, while the promoter of the gene from the ClGPDHg3 genomic clone has little or no activity in the embryo. Thus, for example, a 1387 bp BamHI/AlwNI fragment of ClGPDHg5 is suitable for transcriptional fusion, a 1189 base pair SphI/NarI fragment of ClGPDHg9 is suitable for translational fusion and a 1172 base pair BamHI/BsmAI (part.) fragment of ClGPDHg3 is suitable for transcriptional fusion. Larger (or smaller) promoter fragments can be used for expression of chimeric genes on the basis of additional clones present on the genetic clones. Likewise, any regulatory sequences located downstream from the first codon of the GPDH gene are obtained for targeted expression of chimeric genes from the cloned fragments of genomic DNA.

Northern Blot analysis with polyA⁺-RNA from various *Cuphea lanceolata* tissues with ClGPDH20 cDNA as a probe shows very large amounts of RNA in embryos in comparison with other tissues (see Figure 5). The increase in RNA correlates with increased gene expression and consequently indicates an extremely strong promoter.

The following examples are presented to illustrate this invention.

EXAMPLES

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The plant material used in the context of the present invention was obtained from *Cuphea lanceolata* (Lythraceae) (small lanceolate tube flower).

Example 1

Production of glycerol-3-phosphate dehydrogenase cDNAs

from *Cuphea lanceolata*

A cDNA library was prepared from *Cuphea lanceolata* (wild type) took place with the help of the ZAP[®] cDNA synthesis kit according to the manufacturer's instructions (Stratagene, La Jolla, USA). Messenger RNA from isolated immature embryos about two to three weeks old was used as raw material for the synthesis of the cDNAs. The cDNA library obtained in this way contained 9.5×10^5 recombinant phages.

Functional complementation for isolation of cDNAs that code for a glycerol-3-phosphate dehydrogenase was performed with the *E. Coli* BB26-36 strain (R.M. Bell, J. Bact. 117 (1974) pages 1065-1076). The bacterial medium for culturing BB26-36 (bearing the *plsB26* and *plsX* mutations) was supplemented with 0.1% glycerol to supplement the bacteria. A medium without glycerol was used for functional complementation.

The pBluescript plasmids were cut out of the above cDNA library in 1-ZAP II according to the manufacturer's instructions (Stratagene) by *in vivo* excision using helper phages and then packed in phage coats: 200 ml of XL1Blue *E. Coli* cells ($OD_{600} = 1$) were infected with 5×10^5 pfu of the 1-ZAP II cDNA library, and, in order to guarantee coinfection, were also infected with a tenfold amount of f1 R408 helper phages. After incubating for 15 minutes at a temperature of 37°C for phage adsorption, 5 ml 2xYT medium were added and agitated for three hours more at a temperature of 37°C. During this time, the cells of the pBluescript plasmids packed in the coats of helper phages are secreting the so-called phagemids into the medium. The bacteria were killed

and the 1 phages were inactivated by a heating for 20 minutes at 70°C. After centrifuging, the supernatant containing helper phages along with phagemids was removed. This supernatant was used for infection of the mutant BB26-36 strain.

Complementation was performed after infecting the *E. coli* BB26-36 strain with phagemids containing cDNA plasmids that code for a glycerol-3-phosphate dehydrogenase. M56-LP medium (Bell, loc. cit.) with 50 mg ampicillin was used for selection (without glycerol-3-phosphate). Retransformation of BB26-36 was performed by the method of D. Hanahan, *J. Mol. Biol.* 166 (1983) pages 557-580, with subsequent plating on the selective medium mentioned.

Deletion clones for determining the sequence of the DNA fragments of positive cDNA clones were produced by means of exonuclease III (Strategene) and were sequenced according to the method of Sanger et al., *Proc. Nat. Acad. Sci.* 74 (1977) pages 5463-5467. Some of the DNA sequencing was performed radioactively with the help of the ³²P Sequencing® Kit or with a Pharmacia Automated Laser Fluorescent A.L.F.® DNA sequencer. The sequences were analyzed with the help of computer software from the University of Wisconsin Genetics Computer Group (J. Devereux et al., *Nucl. Acids Res.* 12 (1984) pages 387-394).

Furthermore, cDNA clones were isolated by screening a cDNA library from *Cuphea lanceolata* with CLGPDH20 cDNA as a probe. For this, a cDNA library from *Cuphea lanceolata* (wild type) was produced according to the manufacturer's instructions with the ZAP® cDNA Synthesis Kit. Messenger RNA from isolated, immature embryos about two to three weeks old was the raw material for synthesis of the cDNAs. The cDNA library obtained contained 9.6×10^5 recombinant phages with approx. 50% clones with more than 500 bp insertions. The cDNA library was examined with CLGPDH20 as a probe, and 18 cDNAs were isolated and partially or completely sequenced in the usual manner. Of these cDNAs, 12 were class A, and 6 cDNAs were in class B.

The enzyme measurements were performed with the fusion protein according to the method of Santora et al., Arch. Biochem. Biophys. 196 (1979) pages 403-411.

Example 2

Production of genomic clones of glycerol-3-phosphate dehydrogenase from Cuphea lanceolata

Genomic DNA from young *Cuphea lanceolata* leaves were isolated for this example (S.L. Della Porta et al., Plant. Mol. Biol. Rep. 1, (1983) pages 19-21). The DNA was then partially cleaved with the restriction enzyme Sau3A, whereupon DNA fragments of 11,000 to 19,000 base pairs were cloned in vector lFIXII (Stratagene) that was cleaved with XhoI after the respective interfaces were partially filled with two nucleotides in any given case. The genomic DNA library that was not reproduced amounted to 5.4 times the genome of *Cuphea lanceolata*. Thirty-one genomic clones were then isolated from this library with ClGPDH20-cDNA as a probe.

The three genomic clones ClGPDHg3 (15.9 kb DNA insertion), ClGPDHg5 (17.7 kb DNA insertion) and ClGPDHg9 (15.6 kb DNA insertion) were characterized in greater detail. Suitable subclones were produced in the usual manner and their insertions were sequenced with the ExoIII/Mung bean kit and also with oligonucleotide primers in order to bridge any gaps.

If any of the procedures customary in molecular biology have not have been described adequately here, such procedures were performed by standard methods as described in Sambrook et al., A Laboratory Manual, second edition (1989).

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SEQUENCE LIST

(1) GENERAL INFORMATION:

(i) APPLICANT:

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(E) COUNTRY: Germany

(F) ZIP: 37073

(ii) TITLE OF INVENTION:

Glycerol-3-phosphate dehydrogenase (GPDH)

(iii) NUMBER OF SEQUENCES: 8

(iv) COMPUTER-READABLE FORM:

(A) MEDIUM TYPE: 3.5 inch HD diskette (1.44 MB) /
ASCII Format

(B) COMPUTER: IBM compatible PC

(C) OPERATING SYSTEM: PC-DOS/MS-DOS

(D) SOFTWARE: PatentIn Release #1.0, Version
#1.25 (EPA)

(2) INFORMATION FOR ID SEQ NO:1

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1354 base pairs

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Double strand

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: No

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- (iv) ANTI-SENSE: No
- (vi) ORIGINAL SOURCE:
- (A) ORGANISM: *Cuphea lanceolata*
- (vii) IMMEDIATE SOURCE:
- (A) LIBRARY: ZAP cDNA library
- (B) CLONE: ClGPDH20
- (ix) FEATURE:
- (A) NAME/KEY: cDNA
- (B) LOCATION: 15 to 1345
- (ix) FEATURE:
- (A) NAME/KEY: Fusion with lacZ
- (B) LOCATION: 1 to 14
- (ix) FEATURE:
- (A) NAME/KEY: Start codon
- (B) LOCATION: 17 to 19
- (ix) FEATURE:
- (A) NAME/KEY: Stop codon
- (B) LOCATION: 1133 to 1135
- (ix) FEATURE:
- (A) NAME/KEY: PolyA signal
- (B) LOCATION: 1329 to 1334
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1

GAATTCGGCA CGAGCA ATG GCT CCC TCT GAG CTC AAC TGC ACC CAC CAG	49
Met Ala Pro Ser Glu Leu Asn Cys Thr His Gln	
1 5 10	
AAC CAG CAT TCA AGC GGT TAC GAC GGA CCC AGA TCG AGG GTC ACC GTT	97
Asn Gln His Ser Ser Gly Tyr Asp Gly Pro Arg Ser Arg Val Thr Val	
15 20 25	

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GTC GGT AGT GGA AAC TGG GGT AGT GTT GCT GCC AAG CTC ATT GCT ACC Val Gly Ser Gly Asn Trp Gly Ser Val Ala Ala Lys Leu Ile Ala Thr 30 35 40	145
AAT ACC CTC AAG CTT CCA TCT TTT CAT GAT GAA GTG AGA ATG TGG GTA Asn Thr Leu Lys Leu Pro Ser Phe His Asp Glu Val Arg Met Trp Val 45 50 55	193
TTT GAG GAG ACG CTA CCG AGC GGC GAG AAG CTT ACT GAT GTC ATC AAC Phe Glu Glu Thr Leu Pro Ser Gly Glu Lys Leu Thr Asp Val Ile Asn 60 65 70 75	241
CAG ACC AAT GAA AAT GTT AAG TAT CTC CCC GGA ATT AAG CTC GGT AGG Gln Thr Asn Glu Asn Val Lys Tyr Leu Pro Gly Ile Lys Leu Gly Arg 80 85 90	289
AAT GTT GTT GCA GAT CCA GAC CTC GAA AAC GCA GTT AAG GAT GCA AAT Asu Val Val Ala Asp Pro Asp Leu Glu Asn Ala Val Lys Asp Ala Asn 95 100 105	337
ATG CTC GTG TTT GTG ACA CCG CAT CAG TTC ATG GAG GGC ATC TGC AAA Met Leu Val Phe Val Thr Pro His Gln Phe Met Glu Gly Ile Cys Lys 110 115 120	385
AGA CTC GAA GGG AAA ATA CAA GAA GGA GCA CAG GCT CTC TCC CTT ATA Arg Leu Glu Gly Lys Ile Gln Glu Gly Ala Gln Ala Leu Ser Leu Ile 125 130 135	433
AAG GGC ATG GAG GTC AAA ATG GAG GGG CCT TGC ATG ATC TCG AGC TTA Lys Gly Met Glu Val Lys Met Glu Gly Pro Cys Met Ile Ser Ser Leu 140 145 150 155	481
ATC TCT GAT CTT CTC GGG ATT AAC TGC TGT GTC CTA ATG GGG GCA AAC Ile Ser Asp Leu Leu Gly Ile Asn Cys Cys Val Leu Met Gly Ala Asn 160 165 170	529
ATC GCT AAT GAG ATT GCT GTT GAG AAA TTC AGT GAA GCG ACA GTC GGG Ile Ala Asn Glu Ile Ala Val Glu Lys Phe Ser Glu Ala Thr Val Gly 175 180 185	577
TTC AGA GAA AAT AGA GAT ATT GCA GAG AAA TGG GTT CAG CTC TTT AGC Phe Arg Glu Asn Arg Asp Ile Ala Glu Lys Trp Val Gln Leu Phe Ser 190 195 200	625
ACT CCG TAC TTC ATG GTC TCA GCT GTT GAA GAT GTT GAA GGA GTA GAA Thr Pro Tyr Phe Met Val Ser Ala Val Glu Asp Val Glu Gly Val Glu 205 210 215	673
CTT TGT GGA ACA CTG AAG AAT ATC GTG GCC ATA GCA GCC GGT TTT GTG Leu Cys Gly Thr Leu Lys Asn Ile Val Ala Ile Ala Ala Gly Phe Val 220 225 230 235	721
GAT GGA TTG GAG ATG GGA AAC AAC ACA AAA GCA GCA ATT ATG AGG ATC Asp Gly Leu Glu Met Gly Asn Asn Thr Lys Ala Ala Ile Met Arg Ile	769

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240	245	250	
GGG TTA CGG GAG ATG AAG GCA TTC TCC AAG CTT TTG TTT CCA TCT GTT			817
Gly Leu Arg Glu Met Lys Ala Phe Ser Lys Leu Leu Phe Pro Ser Val			
255	260	265	
AAG GAC ACT ACT TTC TTC GAG AGC TGT GGA GTC GCT GAC CTC ATC ACA			865
Lys Asp Thr Thr Phe Phe Glu Ser Cys Gly Val Ala Asp Leu Ile Thr			
270	275	280	
ACT TGT TTG GGC GGG AGA AAC AGA AAA GTT GCT GAG GCT TTT GCA AAG			913
Thr Cys Leu Gly Gly Arg Asn Arg Lys Val Ala Glu Ala Phe Ala Lys			
285	290	295	
AAT GGC GGG AAA AGG TCA TTC GAT GAT CTC GAA GCA GAG ATG CTC CGG			961
Asn Gly Gly Lys Arg Ser Phe Asp Asp Leu Glu Ala Glu Met Leu Arg			
300	305	310	315
GGG CAA AAA TTA CAG GGT GTC TCA ACA GCA AAG GAG GTC TAT GAA GTC			1009
Gly Gln Lys Leu Gln Gly Val Ser Thr Ala Lys Glu Val Tyr Glu Val			
320	325	330	
TTG GGC CAC CGA GGC TGG CTC GAG CTG TTC CCG CTC TTC TCA ACC GTG			1057
Leu Gly His Arg Gly Trp Leu Glu Leu Phe Pro Leu Phe Ser Thr Val			
335	340	345	
CAC GAG ATA TCC ACT GGC CGT CTG CCT CCT TCA GCC ATC GTC GAA TAC			1105
His Glu Ile Ser Thr Gly Arg Leu Pro Pro Ser Ala Ile Val Glu Tyr			
350	355	360	
AGC GAA CAA AAA ACC ATC TTC TCT TGG TAGAGCAAGA GGCTGCCCTT			1152
Ser Glu Gln Lys Thr Ile Phe Ser Trp			
365	370		
GAAAGACTAA GAGCCACCCT GCCCTGTTTA AAGGGCTAAA AGTTTAATAT TTCTCTGCAG			1212
CCTAAACAGT CGGAAACATT GAAAATCTAG GATGTATAAG AAAAAAAAAA GAAGGTTTGA			1272
AGGAAGTATG GATGGGCATG AATGTATTTA TTTTCGGTAT ACTCTTTTTC TGCAAAAATA			1332
ATTTCTTCAG AAAGGGGGGC CC			1354

(2) INFORMATION FOR ID SEQ NO:2

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1464 base pairs
- (B) TYPE: Nucleic acid
- (C) STRANDEDNESS: Double stranded
- (D) TOPOLOGY: Linear

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(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: No

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Cuphea lanceolata

(vii) IMMEDIATE SOURCE:

(A) LIBRARY: ZAP cDNA library

(B) CLONE: ClGPDH109

(ix) FEATURE:

(A) NAME/KEY: cDNA

(B) LOCATION: 15 to 1454

(ix) FEATURE:

(A) NAME/KEY: CDS [coding sequence]

(B) LOCATION: 15 to 1187

(ix) FEATURE:

(A) NAME/KEY: Fusion with lacZ

(B) LOCATION: 1 to 14

(ix) FEATURE:

(A) NAME/KEY: Start codon

(B) LOCATION: 45 to 47

(ix) FEATURE:

(A) NAME/KEY: Stop codon

(B) LOCATION: 1188 to 1190

(ix) FEATURE:

(A) NAME/KEY: PolyA signal

(B) LOCATION: 1414 to 1419

(ix) FEATURE:

2170611

(A) NAME/KEY: PolyA region

(B) LOCATION: 1446 to 1454

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2

GAATTCGGCA CGAGCTTCCT CTGTTCTTCC TCTCTGCCTC TGCA ATG GCG CCT GCC	56
Met Ala Pro Ala	
1	
TTC GAA CCC CAT CAG CTG GCT CCC TCT GAG CTT AAC TCT GCC CAC CAG	104
Phe Glu Pro His Gln Leu Ala Pro Ser Glu Leu Asn Ser Ala His Gln	
5 10 15 20	
AAC CCA CAT TCA GGC GGA TAT GAC GGA CCC AGA TCG AGG GTC ACT GTC	152
Asn Pro His Ser Gly Gly.Tyr Asp Gly Pro Arg Ser Arg Val Thr Val	
25 30 35	
GTC GGC AGC GGC AAC TGG GGC AGC GTC GCT GCC AAG CTC ATT GCT TCC	200
Val Gly Ser Gly Asn Trp Gly Ser Val Ala Ala Lys Leu Ile Ala Ser	
40 45 50	
AAC ACC CTC AAG CTC CCA TCT TTC CAT GAT GAA GTG AGG ATG TGG GTA	248
Asu Thr Leu Lys Leu Pro Ser Phe His Asp Glu Val Arg Met Trp Val	
55 60 65	
TTT GAG GAG ACT CTA CCG GGC GGC GAG AAG CTC ACT GAT ATC ATC AAC	296
Phe Glu Glu Thr Leu Pro Gly Gly Glu Lys Leu Thr Asp Ile Ile Asn	
70 75 80	
CAG ACC AAT GAA AAT GTT AAA TAT CTT CCC GGA ATT AAG CTC GGT GGG	344
Glu Thr Asn Glu Asn Val Lys Tyr Leu Pro Gly Ile Lys Leu Gly Gly	
85 90 95 100	
AAT GTT GTT GCT GAT CCA GAC CTC GAA AAT GCA GTT AAG GAT GCA AAT	392
Asn Val Val Ala Asp Pro Asp Leu Glu Asn Ala Val Lys Asp Ala Asn	
105 110 115	
ATG CTC GTG TTT GTC ACA CCG CAT CAG TTC ATG GAG GGC ATC TGC AAA	440
Met Leu Val Phe Val Thr Pro His Gln Phe Met Glu Gly Ile Cys Lys	
120 125 130	
AGA CTT GTC GGG AAG ATA CAG GAA GGA GCG CAG GCT CTC TCC CTT ATA	488
Arg Leu Val Gly Lys Ile Gln Glu Gly Ala Gln Ala Leu Ser Leu Ile	
135 140 145	
AAA GGC ATG GAG GTC AAG ATG GAG GGG CCT TGC ATG ATC TCG AGC CTA	536
Lys Gly Met Glu Val Lys Met Glu Gly Pro Cys Met Ile Ser Ser Leu	
150 155 160	
ATC TCA GAT CTT CTC GGG ATC AAC TGC TGT GTC CTT AAT GGG GCA AAC	584
Ile Ser Asp Leu Leu Gly Ile Asu Cys Cys Val Leu Asn Gly Ala Asn	

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165	170	175	180	
ATC GCT AAT GAG ATT GCT GTT GAG AAA TTC AGT GAA GCG ACT GTC GGG				632
Ile Ala Asn Glu Ile Ala Val Glu Lys Phe Ser Glu Ala Thr Val Gly	185	190	195	
TTC AGA GAA AAT AGA GAT ATT GCG GAA AAA TGG GTT CAG CTC TTT AGC				680
Phe Arg Glu Asn Arg Asp Ile Ala Glu Lys Trp Val Gln Leu Phe Ser	200	205	210	
ACT CCA TAC TTC ATG GTC TCA GCT GTT GAA GAT GTT GAA GGA GTA GAG				728
Thr Pro Tyr Phe Met Val Ser Ala Val Glu Asp Val Glu Gly Val Glu	215	220	225	
CTT TGT GGA ACA CTG AAG AAT ATT GTG GCC ATA GCA GCG GGT TTT GTT				776
Leu Cys Gly Thr Leu Lys Asn Ile Val Ala Ile Ala Ala Gly Phe Val	230	235	240	
GAT GGA TTG GAG ATG GGA AAC AAC ACA AAA GCG GCA ATT ATG AGG ATC				824
Asp Gly Leu Glu Met Gly Asn Asn Thr Lys Ala Ala Ile Met Arg Ile	245	250	255	260
GGG CTG CGG GAG ATG AAA GCG TTC TCC AAG CTT TTG TTT CCA TCT GTT				872
Gly Leu Arg Glu Met Lys Ala Phe Ser Lys Leu Leu Phe Pro Ser Val	265	270	275	
AAG GAC ACT ACT TTT TTC GAG AGC TGC GGA GTC GCT GAT CTC ATC ACA				920
Lys Asp Thr Thr Phe Phe Glu Ser Cys Gly Val Ala Asp Leu Ile Thr	280	285	290	
ACT TGT TTG GGC GGA AGA AAC AGA AAA GTC GCT GAG GCT TTT GCA AAG				968
Thr Cys Leu Gly Gly Arg Asn Arg Lys Val Ala Glu Ala Phe Ala Lys	295	300	305	
AAT GGC GGA AAC AGG TCA TTT GAT GAT CTC GAA GCA GAG ATG CTC CGG				1016
Asn Gly Gly Asn Arg Ser Phe Asp Asp Leu Glu Ala Glu Met Leu Arg	310	315	320	
GGG CAA AAA TTA CAG GGT GTC TCG ACA GCG AAA GAG GTC TAC GAG GTC				1064
Gly Gln Lys Leu Gln Gly Val Ser Thr Ala Lys Glu Val Tyr Glu Val	325	330	335	340
CTG AGG CAC CGA GGC TGG CTC GAG TTG TTC CCG CTC TTC TCA ACC GTG				1112
Leu Arg His Arg Gly Trp Leu Glu Leu Phe Pro Leu Phe Ser Thr Val	345	350	355	
CAT GAG ATC TCC AGT GGC CGT CTG CCT CCT TCA GCC ATT GTT GAA TAC				1160
His Glu Ile Ser Ser Gly Arg Leu Pro Pro Ser Ala Ile Val Glu Tyr	360	365	370	
AGC GAA CAA AAG CCT ACC TTC TCT TGG TAGAGAAAGA AACCAGGAAG				1207
Ser Glu Gln Lys Pro Thr Phe Ser Trp	375	380		

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AACGGCGAGC CACTGTCCCC CGTTTAAAGG TTTACTATTT CTCTCTGCAC TTTGCAGCCT 1267
GAAGAGTCGG AACATAGAA AATCTAGGAA GTTTCAGAAA AAGGAAGGTT TTGAGGATGT 1327
ATGGATGATA TATATACTAG GTGGGTATGA AGAGGAAGTT ATTACTATGA TGTGTTGATG 1387
TGGTAATGGC TAAGTACATG AGATCAAATA AATAGACAGA CCTTGGTTTC TTCTTTCTAA 1447
AAAAAAAGGG GGGGCCC 1464

(2) INFORMATION FOR ID SEQ NO:3

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1490 base pairs

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Double

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: No

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Cuphea lanceolata*

(vii) IMMEDIATE SOURCE:

(A) LIBRARY: ZAP cDNA library

(B) CLONE: ClGPDH132

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 15 to 1115

(ix) FEATURE:

(A) NAME/KEY: Fusion with lacZ

(B) LOCATION: 1 to 14

(ix) FEATURE:

(A) NAME/KEY: Stop codon

(B) LOCATION: 1116 to 1118

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(ix)

FEATURE:

(A) NAME/KEY: PolyA signal

(B) LOCATION: 1343 to 1348

(ix)

FEATURE:

(A) NAME/KEY: PolyA region

(B) LOCATION: 1465 to 1484

(xi)

SEQUENCE DESCRIPTION: SEQ ID NO:3

GAATTCGGCA CGAG CTT AAC TCT GCC CAC CAG AAC CCA CAT TCC AGC GGA	50
Leu Asn Ser Ala His Gin Asn Pro His Ser Ser Gly	
1 5 10	
TAT GAA GGA CCC AGA TCG AGG GTC ACC GTC GTT GGC AGC GGC AAC TGG	98
Tyr Glu Gly Pro Arg Ser Arg Val Thr Val Val Gly Ser Gly Asn Trp	
15 20 25	
GGC AGC GTC GCT GCC AAG CTC ATT GCT TCC AAC ACC CTC AAG CTC CCA	146
Gly Ser Val Ala Ala Lys Leu Ile Ala Ser Asn Thr Leu Lys Leu Pro	
30 35 40	
TCT TTC CAT GAT GAA GTG AGG ATG TGG GTA TTT GAG GAG ACT CTA CCG	194
Ser Phe His Asp Glu Val Arg Met Trp Val Phe Glu Glu Thr Leu Pro	
45 50 55 60	
GGC GGC GAG AAG CTC ACT GAT ATC ATC AAC CAG ACC AAT GAA AAT GTT	242
Gly Gly Glu Lys Leu Thr Asp Ile Ile Asn Gln Thr Asn Glu Asn Val	
65 70 75	
AAA TAT CTT CCC GGA ATT AAG CTC GGT AGG AAT GTT GTT GCA GAT CCA	290
Lys Tyr Leu Pro Gly Ile Lys Leu Gly Arg Asn Val Val Ala Asp Pro	
80 85 90	
GAC CTC GAA AAC GCA GTT AAG GAT GCA AAT ATG CTC GTT TTC GTC ACA	338
Asp Leu Glu Asn Ala Val Lys Asp Ala Asn Met Lou Val Phe Val Thr	
95 100 105	
CCG CAT CAG TTC GTG GAG GGC ATC TGC AAA AGA CTT GTA GGG AAG ATA	386
Pro His Gin Phe Val Glu Gly Ile Cys Lys Arg Leu Val Gly Lys Ile	
110 115 120	
CAG GAA GGA GCG CAG GCT CTC TCT CTT ATA AAA GGC ATG GAG GTC AAA	434
Gin Glu Gly Ala Gin Ala Leu Ser Leu Ile Lys Gly Met Glu Val Lys	
125 130 135 140	
ATG GAG GGG CCT TGC ATG ATC TCG AGC CTA ATC TCA GAT CTT CTC GGG	482
Met Glu Gly Pro Cys Met Ile Ser Ser Leu Ile Ser Asp Leu Leu Gly	
145 150 155	

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ATC AAT TGC TGT GTC CTT AAT GGG GCG AAC ATC GCT AAT GAG ATT GCT Ile Asn Cys Cys Val Leu Asn Gly Ala Asn Ile Ala Asn Glu Ile Ala 160 165 170	530
GTT GAG AAA TTC AGT GAA GCG ACT GTC GGG TTC AGA GAA AAT AGA GAT Val Glu Lys Phe Ser Glu Ala Thr Val Gly Phe Arg Glu Asn Arg Asp 175 180 185	578
ATT GCG GAA AAA TGG GTT CAG CTC TTT AGC ACT CCA TAC TTC ATG GTC Ile Ala Glu Lys Trp Val Gln Leu Phe Ser Thr Pro Tyr Phe Met Val 190 195 200	626
TCA GCT GTT GAA GAT GTT GAA GGA GTA GAG CTT TGT GGA ACA CTG AAG Ser Ala Val Glu Asp Val Glu Gly Val Glu Leu Cys Gly Thr Leu Lys 205 210 215 220	674
AAT ATT GTG GCC ATA GCA GCG GGT TTT GTG GAT GGA CTG GAG ATG GGA Asu Ile Val Ala Ile Ala Ala Gly Phe Val Asp Gly Leu Glu Met Gly 225 230 235	722
AAC AAC ACA AAA GCA GCA ATT ATG AGG ATC GGG CTG CGG GAG ATG AAA Asn Asn Thr Lys Ala Ala Ile Met Arg Ile Gly Leu Arg Glu Met Lys 240 245 250	770
GCG TTC TCC AAG CTT TTG TTT CCA TCT GTT AAG GAC ACT ACT TTT TTC Ala Phe Ser Lys Leu Leu Phe Pro Ser Val Lys Asp Thr Thr Phe Phe 255 260 265	818
GAG AGC TGC GGA GTC GCT GAT CTC ATC ACA ACT TGT TTG GGC GGA AGA Glu Ser Cys Gly Val Ala Asp Leu Ile Thr Thr Cys Leu Gly Gly Arg 270 275 280	866
AAC AGA AAA GTC GCT GAG GCT TTT GCA AAG AAT GGC GGT AAC AGG TCA Asn Arg Lys Val Ala Glu Ala Phe Ala Lys Asn Gly Gly Asn Arg Ser 285 290 295 300	914
TTC GAT GAT CTC GAA GCA GAG ATG CTC CGG GGG CAA AAA TTA CAG GGT Phe Asp Asp Leu Glu Ala Glu Met Leu Arg Gly Gln Lys Leu Gln Gly 305 310 315	962
GTC TCG ACA GCG AAA GAG GTC TAC GAG GTC CTG AGG CAC CGA GGT TGG Val Ser Thr Ala Lys Glu Val Tyr Glu Val Leu Arg His Arg Gly Trp 320 325 330	1010
CTC GAG TTG TTC CCG CTC TTC TCA ACC GTG CAT GAG ATC TCC ACT GGC Leu Glu Leu Phe Pro Leu Phe Ser Thr Val His Glu Ile Ser Thr Gly 335 340 345	1058
CGT CTG CCT CCT TCA GCC ATT GTT GAA TAC AGC GAA CAA AAG CCC ACC Arg Leu Pro Pro Ser Ala Ile Val Glu Tyr Ser Glu Gln Lys Pro Thr 350 355 360	1106
TTC TCT TGG TAGAGAAAGA AGCAACCAGG AAGAACGGCG AGCCACTCTG Phe Ser Trp	1155

365

CCTCGTTTAA AGGGTTACTA TTTCTCTACA CTCTGCAGCC TGAAGAGTCG GAAACATCGA 1215
 AAATCTAGGA AGTCTCAGAA AAATGAAGGT TTGGAGGATG TATGGATGAT ATATATACTA 1275
 GGTGGGTATG AAGAGGAAGT TATTACTATG ATGTTGGTAT GTGGTAATGG CTAAGTACAT 1335
 GAGATCAAAT AAATAGACAG ACCTTGGTTT CTTCTATCTC GATTCGGTCT CGTCGAGTTT 1395
 GCGGAAACTC AACTGAACTT CCTGAGTACC CTGCTACCTA TTACATGTAA TGTTCTTATT 1455
 TATATGCTTA AAAAAAAAAA AAAAAAAAAAC TCGAG 1490

(2) INFORMATION FOR ID SEQ NO:4

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1390 base pairs
 - (B) TYPE: Nucleic acid
 - (C) STRANDEDNESS: Double strand
 - (D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: No
- (iv) ANTI-SENSE: No
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: *Cuphea lanceolata*
- (vii) IMMEDIATE SOURCE:
 - (A) LIBRARY: ZAP cDNA library
 - (B) CLONE: C1GPDH30
- (ix) FEATURE:
 - (A) NAME/KEY: cDNA
 - (B) LOCATION: 15 to 1384
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 34 to 1149
- (ix) FEATURE:
 - (A) NAME/KEY: Fusion with lacZ
 - (B) LOCATION: 1 to 14

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- (ix) FEATURE:
- (A) NAME/KEY: Start codon
- (B) LOCATION: 34 to 36
- (ix) FEATURE:
- (A) NAME/KEY: Stop codon
- (B) LOCATION: 1150 to 1152
- (ix) FEATURE:
- (A) NAME/KEY: PolyA signal
- (B) LOCATION: 1349 to 1354
- (ix) FEATURE:
- (A) NAME/KEY: PolyA region
- (B) LOCATION: 1366 to 1384
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4

GAATTCGGCA CGAGTTTCTT CTCAGCCTCT GCA ATG GCT CCC TCT GAG CTC AAC	54
Met Ala Pro Ser Glu Leu Asn	
1 5	
TGC ACC CAC CAG AAC CCA CAT TCA AGC GGT TAC GAC GGA CCC AGA TCG	102
Cys Thr His Gln Asn Pro His Ser Ser Gly Tyr Asp Gly Pro Arg Ser	
10 15 20	
AGG GTC ACC GTT GTC GGT AGT GGA AAC TGG GGC AGT GTC GCT GCC AAG	150
Arg Val Thr Val Val Gly Ser Gly Asn Trp Gly Ser Val Ala Ala Lys	
25 30 35	
CTC ATT GCT TCC AAT ACC CTC AAG CTT CCA TCT TTT CAT GAT GAA	
Leu Ile Ala Ser Asn Thr Leu Lys Leu Pro Ser Phe His Asp Glu	
40 45 50	
AGA ATG TGG GTA TTT GAG GAG ACT CTA CCG AGC GGC GAG AAG CTT ACT	
Arg Met Trp Val Phe Glu Glu Thr Leu Pro Ser Gly Glu Lys Leu Thr	
60 65 70	
GAT GTC ATC AAC CAG ACC AAT GAA AAT GTT AAG TAT CTC CCC GGA ATT	294
Asp Val Ile Asn Gln Thr Asn Glu Asn Val Lys Tyr Leu Pro Gly Ile	
75 80 85	
AAG CTC GGT AGG AAT GTT GTT GCA GAT CCA GAC CTC GAA AAC GCA GTT	342
Lys Leu Gly Arg Asn Val Val Ala Asp Pro Asp Leu Glu Asn Ala Val	

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90	95	100	
AAG GAT GCA AAT ATG CTC GTG TTT GTG ACA CCG CAT CAG TTC ATG GAG Lys Asp Ala Asn Met Leu Val Phe Val Thr Pro His Gln Phe Met Glu 105 110 115			390
GGC ATC TGC AAA AGA CTC GTA GGG AAA ATA CAG GAA GGA GCA CAG GCT Gly Ile Cys Lys Arg Leu Val Gly Lys Ile Gln Glu Gly Ala Gln Ala 120 125 130 135			438
CTC TCC CTT ATA AAG GGC ATG GAG GTC AAA ATG GAG GGG CCT TGC ATG Leu Ser Leu Ile Lys Gly Met Glu Val Lys Met Glu Gly Pro Cys Met 140 145 150			486
ATC TCG AGC CTA ATC TCT GAT CTT CTC GGG ATC AAC TGC TGT GTC CTA Ile Ser Ser Leu Ile Ser Asp Leu Leu Gly Ile Asn Cys Cys Val Leu 155 160 165			534
ATG GGG GCA AAC ATC GCT AAT GAG ATT GCT GTT GAG AAA TTC AGT GAA Met Gly Ala Asn Ile Ala Asn Glu Ile Ala Val Glu Lys Phe Ser Glu 170 175 180			582
GCG ACA GTC GGG TTC AGA GAA AAT ACA GAT ATT GCG GAG AAA TGG GTT Ala Thr Val Gly Phe Arg Glu Asn Thr Asp Ile Ala Glu Lys Trp Val 185 190 195			630
CAG CTC TTT AGC ACT CCG TAC TTC ATG GTC TCA GCT GTT GAA GAT GTT Gln Leu Phe Ser Thr Pro Tyr Phe Met Val Ser Ala Val Glu Asp Val 200 205 210 215			678
GAA GGA GTA GAA CTT TGT GGA ACA CTG AAG AAT ATC GTG GCC ATA GCA Glu Gly Val Glu Leu Cys Gly Thr Leu Lys Asn Ile Val Ala Ile Ala 220 225 230			726
GCC GGT TTT GTG GAT GGA TTG GAG ATG GGA AAC AAC ACA AAA GCA GCA Ala Gly Phe Val Asp Gly Leu Glu Met Gly Asn Asn Thr Lys Ala Ala 235 240 245			774
ATT ATG AGG ATC GGG TTA CGG GAG ATG AAG GCA TTC TCC AAG CTT TTG Ile Met Arg Ile Gly Leu Arg Glu Met Lys Ala Phe Ser Lys Leu Leu 250 255 260			822
TTT CCA TCT GTT AAG GAC ACT ACT TTC TTC GAG AGC TGT GGA GTT GCT Phe Pro Ser Val Lys Asp Thr Thr Phe Phe Glu Ser Cys Gly Val Ala 265 270 275			870
GAC CTC ATC ACA ACT TGT TTG GGC GGG AGA AAC AGA AAA GTT GCT GAG Asp Leu Ile Thr Thr Cys Leu Gly Gly Arg Asn Arg Lys Val Ala Glu 280 285 290 295			918
GCT TTT GCA AAG KAT GGC GGG GAA AGG TCA TTC GAT GAT CTC GAA GCA Ala Phe Ala Lys Asn Gly Gly Glu Arg Ser Phe Asp Asp Leu Glu Ala 300 305 310			966

GAG CTG CTC CGG GGG CAA AAA TTA CAG GGT GTC TCA ACA GCA AAG GAG Glu Leu Leu Arg Gly Gln Lys Leu Gln Gly Val Ser Thr Ala Lys Glu 315 320 325	1014
GTC TAT GAA GTC TTG GGG CAC CGA GGC TGG CTC GAG CTG TTC CCG CTC Val Tyr Glu Val Leu Gly His Arg Gly Trp Leu Glu Leu Phe Pro Leu 330 335 340	1062
TTC TCA ACC GTG CAC GAG ATC TCC ACT GGC CGT CTG CAT CCT TCA GCC Phe Ser Thr Val His Glu Ile Ser Thr Gly Arg Leu His Pro Ser Ala 345 350 355	1110
ATC GTC GAA TAC AGC GAA CAA AAA ACC ATC TTC TCT TGG TAGAGCAAGA Ile Val Glu Tyr Ser Glu Gln Lys Thr Ile Phe Ser Trp 360 365 370	1159
GGCTGCCCTT GAAAGACTAA GAGCCACCCT GCCCTGTTTA AAGGGCTAAA AGTTTAATAT TTCTCTGCAG CCTAAACAGT TGGAAACATT GAAAATCTAG GATGTATCAG AAAAAAGAAG GTTTGGAGGA AGTATGGATG ATATAGAGGA CATGAATGTA TTCATTTTCG GTATACTCTT TTTCTGCAAA ATAATTCTTC AGATGTAAAA AAAAAAAAAA AAAAACTCGA G	1219 1279 1339 1390

(2) INFORMATION FOR ID SEQ NO:5

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4434 base pairs
- (B) TYPE: Nucleic acid
- (C) STRANDEDNESS: Double strand
- (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: No

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Cuphea lanceolata*

(vii) IMMEDIATE SOURCE:

(A) LIBRARY: Genomic lambda FIX II

(B) CLONE: ClGPDH5

(ix) FEATURE:

(A) NAME/KEY: TATA signal

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(B) LOCATION: 1332 to 1336

(ix) FEATURE:

(A) NAME/KEY: Start codon

(B) LOCATION: 1394 to 1396

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: Join (1394 to 1550, 2066 to 2142, 2241 to 2313, 2405 to 2622, 2719 to 2826, 2961 to 3024, 3233 to 3260, 3342 to 3462, 3541 to 3595, 3692 to 3740, 3580 to 4005)

(ix) FEATURE:

(A) NAME/KEY: Stop codon

(B) LOCATION: 4006 to 4008

(ix) FEATURE:

(A) NAME/KEY: PolyA signal

(B) LOCATION: 4205 to 4210

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5

GGATCCTTAG	AAGACAAGCG	CGGGGCGGGC	ATGGGTCTCG	TGATACCCGC	CCCATTTTGC	60
CCCATTCAT	CCCTATATGG	TAAGCAGATC	TCACTGAAAA	GTCACCGTTT	CTGGATGGTT	120
TCCAGATGAT	TTTGTCCCTC	CCTCTAGCTG	CATTAGGTGA	TGGGATTGAG	GCTATTCTAA	180
GAACCAGCTC	GTGTGGAAGG	TAGGCGGAGA	TTAGCTCCCA	GTTCCATCCT	CCTGTATTTG	240
AAGCGAAGAA	AGAAACTGGG	TTGTCTAGCA	TGTTTTGTGG	GACAGGTTTG	GTCGTCTTTT	300
CTGATAGGCT	CTGATTCAAT	AGAAGCCAAT	TATCTCTCCA	AAAGGAAACC	TTATTACCAC	360
TTCCAATCGA	CCACCCTATG	TACTTGCTGA	TCTTCGGCCA	GGTATCGCAT	AAAGCATTCC	420
ATAACGCTGA	TGCTGTCGTC	TTTTTTGTGA	ATGTTGGCAA	GAGTGTGTCT	GGCATGGCAT	480
ATTTGTGACT	GAGCACCCGC	ACCCAAAGGC	TCTGAGGTTG	TGATGCCATA	TCCCAACATA	540
CCTTCGATAG	AAAGGCTTCA	TTCATCTTCC	GTAGCTTACG	AATGCCAAGA	CCACCCCATG	600
GTGCTGGACT	AGTGACCGTG	GACCAATTGA	CCAAATGCAC	CTTCCTTTGC	TCCATTGAAT	660
GGCCCCAAAT	GAAGTTGCCG	CAATGTCTTT	CGATTTTCATC	AAGTGTTCCT	TGAGGAATAC	720
GTGTGGACTG	CATGGAGAAG	GATGGCAGAG	CCGTCAAGAC	AGATTTACCC	AGCGTCACCC	780
GCCCAGCCAT	TGACAGTGTC	GATGCCGACC	AACCAGCAAG	TCTTGCTTTT	ACCTCGACAT	840
GTTTTGGATT	TTATATACCG	GTGGTGATGG	TGTTTGAATT	AATCATCGTC	ATTAATTTAT	900
ACCGTGCAAT	ATATATTGCA	ACATTCCAAA	GTATAATTAA	TTTTATATGT	CCATTCGTGA	960
CTAATCTTGG	AGATAGGGCT	TAAATTGTTA	TATGATGATA	TAGAAGAAGT	TGGATAGCAC	1020
ATAAGAACTC	TATAAAATGC	TTATAGATCA	TGGCATCGAA	TTCATCCGCT	ATATATGAGT	1080

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GAGGAAGAAA CTAATCAAAA CCTCGTATTC ATCGAAACAA CCGTTGAAGT GGTTACACTT	1140
TGAATCCTAA GACATACTTG ACGTCATGAT TCTGTCTCTC TATTCCATTG CATAATAAAT	1200
AAAACAAAGG AAACAAAAGC ATAGAGGAGA TCGCCAGATT CAGCAGTTTC CGCATAGGTT	1260
GCCACGGAGC CTTACATGCC GATGCCTTCC TCTGCCTCCT TCTTCCTCCT GTCTCTCTCT	1320
CTACATCCCC TTATATCCCT TCCTCCTTCC CTCCATCTTC ACCATTCTC TGTTTTTCTT	1380
CTCAGCCTCT GCA ATG GCT CCC TCT GAG CTC AAC TGC ACC CAC CAG AAC	1429
Met Ala Pro Ser Glu Leu Asn Cys Thr His Gln Asn	
1 5 10	
CCA CAT TCA AGC GGT TAC GAC GGA CCC AGA TCG AGG GTC ACC GTT GTC	1477
Pro His Ser Ser Gly Tyr Asp Gly Pro Arg Ser Arg Val Thr Val Val	
15 20 25	
GGT AGT GGA AAC TGG GGC AGT GTC GCT GCC AAG CTC ATT GCT TCC AAT	1525
Gly Ser Gly Asn Trp Gly Ser Val Ala Ala Lys Leu Ile Ala Ser Asn	
30 35 40	
ACC CTC AAG CTT CCA TCT TTT CAT G GTTCGTCTCT CCTTTTCTCT	1570
Thr Leu Lys Leu Pro Ser Phe His	
45 50	
GAAAAATGAA GCTTTTGCAT GGGATAGTCA CTAGATATGA GCCTCTGTTT GCATGACTGA	1630
AGCGCTTGAG TAACCGAGTT TTTGGAACAA GAGCACAGGT GGTTCCTTTG CATTTCCTTT	1690
GAGGTTCTTT AATCATTCAA TGAAGTAGCG GTTGATCGCT GAGCAATTGA AACTTGTGGA	1750
ATCGAACCTC CAGCCGAGTC TTAGTGTAAT TGCTTTCTGT TTTACTTCAT TCATAGTGGG	1810
AAGGAGTACG AACTGATGAG TGATGTCACA TTTCATTAGT CGGGTGCGA AAAAATCAG	1870
TTGACATATT GGTGAGACT CTGCAGTGTC ATCAGATATG AGTTGGTGTA TTTGTATTGA	1930
CATTGGAATT TGGTATGTGT ATGAATTTTG TTGAATTAAT CACCGCTGTG ATGAAAAGAT	1990
CAGTACTTCT TCGGTCATTT TTCAGGTGGA AGGATGTTGG TTTCTTATAT ATGTAACCTT	2050
ACATGAATTT TTCAG AT GAA GTG AGA ATG TGG GTA TTT GAG GAG ACT CTA	2100
Asp Glu Val Arg Met Trp Val Phe Glu Glu Thr Leu	
55 60	
CCG AGC GGC GAG AAG CTT ACT GAT GTC ATC AAC CAG ACC AAT	2142
Pro Ser Gly Glu Lys Leu Thr Asp Val Ile Asn Gln Thr Asn	
65 70 75	
GTAAGGAAAC ACAGATTAGC AATAGCATGA GCAGTTATTG CTGGTTAAAT ATGCTTGTTA	2202
GCAACTTTTCG TGACGGCCTG AGTTTTATAC CTCTGCAG GAA AAT GTT AAG TAT	2255
Glu Asn Val Lys Tyr	
80	
CTC CCC GGA ATT AAG CTC GGT AGG AAT GTT GTT GCA GAT CCA GAC CTC	2303
Leu Pro Gly Ile Lys Leu Gly Arg Asn Val Val Ala Asp Pro Asp Leu	
85 90 95	
GAA AAC GCA G GTAGTCCATG TGTTCAATTAG AATTCTCTAA TTAATTATTG	2353
Glu Asn Ala	
100	
TGGTTTATTT CCTTGTCTCT GTGATGATAT TCTGGATGAA ATTTTGTGCA G TT AAG	2409
Val Lys	

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GAT GCA AAT ATG CTC GTG TTT GTG ACA CCG CAT CAG TTC ATG GAG GGC Asp Ala Asn Met Leu Val Phe Val Thr Pro His Gln Phe Met Glu Gly 105 110 115 120	2457
ATC TGC AAA AGA CTC GTA GGG AAA ATA CAG GAA GGA GCA CAG GCT CTC Ile Cys Lys Arg Leu Val Gly Lys Ile Gln Glu Gly Ala Gln Ala Leu 125 130 135	2505
TCC CTT ATA AAG GGC ATG GAG GTC AAA ATG GAG GGG CCT TGC ATG ATC Ser Leu Ile Lys Gly Met Glu Val Lys Met Glu Gly Pro Cys Met Ile 140 145 150	2553
TCG AGC CTA ATC TCT GAT CTT CTC GGG ATC AAC TGC TGT GTC CTA ATG Ser Ser Leu Ile Ser Asp Leu Leu Gly Ile Asn Cys Cys Val Leu Met 155 160 165	2601
GGG GCA AAC ATC GCT AAT GAG GTAAACACTT GGCACGATCT GGTGCAACT Gly Ala Asn Ile Ala Asn Glu 170 175	2652
CCCCCAGGAA ATTGTAGATC CTCATACTGT TAGCATCTTG ATGAGGTTAA ATATCTTATG TTGTAG ATT GCT GTT GAG AAA TTC AGT GAA GCG ACA GTC GGG TTC AGA Ile Ala Val Glu Lys Phe Ser Glu Ala Thr Val Gly Phe Arg 180 185	2712 2760
GAA AAT ACA GAT ATT GCG GAG AAA TGG GTT CAG CTC TTT AGC ACT CCG Glu Asn Thr Asp Ile Ala Glu Lys Trp Val Gln Leu Phe Ser Thr Pro 190 195 200 205	2808
TAC TTC ATG GTC TCA GCT GTAAGTTGCG ATAAACCTT ACGTTTTGCT Tyr Phe Met Val Ser Ala 210	2856
AATAGAACAC AATGCTAGAA ACTCCCAGAT TTCAATGTTA TGTATTTTGG TGCCCAAAGA AGCAACTTCT TAACATCTGT GGCTCCTCTT ACTGACAAAA ATAG GTT GAA GAT GTT Val Glu Asp Val 215	2916 2972
GAA GGA GTA GAA CTT TGT GGA ACA CTG AAG AAT ATC GTG GCC ATA GCA Glu Gly Val Glu Leu Cys Gly Thr Leu Lys Asn Ile Val Ala Ile Ala 220 225 230	3020
GCC G GTTCGTGTTT ACGAGATGTA CATTTATGTA TAACAATCTT TCATTTATTC Ala	3074
ATCGAGATGG GATGCAATAT ATCAATGAGA GGGAAAAGAA AGGGCAAAGG AAAATGCTGT	3134
TGTATTGCAG CTTTAGGCAT TCTTTTCTCT TAATTATTAA CTGTGAAACA CCGAGAAGTA	3194
TTGATGAAGT TAAGAAACGA TGTTACAG GT TTT GTG GAT GGA TTG GAG ATG	3245

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	Gly Phe Val Asp Gly Leu Glu Met	
	235	240
GGA AAC AAC ACA AAA GTAAGTCTAA ATTTTGTGTA AAACCTTAAAG TAAGAGTTTA		3300
Gly Asn Asn Thr Lys		
245		
TGCTTTGGCA TTGTTTGAAG TTCACCTTACT AATGACTTTA G GCA GCA ATT ATG		3353
Ala Ala Ile Met		
AGG ATC GGG TTA CGG GAG ATG AAG GCA TTC TCC AAG CTT TTG TTT CCA		3401
Arg Ile Gly Leu Arg Glu Met Lys Ala Phe Ser Lys Leu Leu Phe Pro		
250 255 260 265		
TCT GTT AAG GAC ACT ACT TTC TTC GAG AGC TGT GGA GTT GCT GAC CTC		3449
Ser Val Lys Asp Thr Thr Phe Phe Glu Ser Cys Gly Val Ala Asp Leu		
270 275 280		
ATC ACA ACT TGT T GTAAGGAAGC ATATAGATTT CCTTCGAATA TGAATAAATT		3502
Ile Thr Thr Cys		
285		
GCATAGTTCA TATCATCATA ATTTGTGTTT GTGCTCAG TG GGC GGG AGA AAC		3554
Leu Gly Gly Arg Asn		
290		
AGA AAA GTT GCT GAG GCT TTT GCA AAG AAT GGC GGG GAA AG		3595
Arg Lys Val Ala Glu Ala Phe Ala Lys Asn Gly Gly Glu Arg		
295 300		
GTCGTGTTTC CCTTTCGTCG ATCCTGATTT AATTCCTGTT TAGTGGTATT CACTTTGTGT		3655
GTATGTAAAT CAAGCAACTA TTTCCATCAT CTTCAG G TCA TTC GAT GAT CTC		3707
Ser Phe Asp Asp Leu		
305		
GAA GCA GAG CTG CTC CGG GGG CAA AAA TTA CAG GTACATGATG AAGAAACCGA		3760
Glu Ala Glu Leu Leu Arg Gly Gln Lys Leu Gln		
310 315 320		
TGTCTATACA GAAAGAGTCC ATTGCAAAGC TTGAGAATGT TTCGAGCATA AAGAGCATAA		3820
GAATATTCTT TTCGGTGATT TTCATGCAG GGT GTC TCA ACA GCA AAG GAG GTC		3873
Gly Val Ser Thr Ala Lys Glu Val		
325		
TAT GAA GTC TTG GGG CAC CGA GGC TGG CTC GAG CTG TTC CCG CTC TTC		3921
Tyr Glu Val Leu Gly His Arg Gly Trp Leu Glu Leu Phe Pro Leu Phe		
330 335 340		
TCA ACC GTG CAC GAG ATC TCC ACT GGC CGT CTG CAT CCT TCA GCC ATC		3969
Ser Thr Val His Glu Ile Ser Thr Gly Arg Leu His Pro Ser Ala Ile		
345 350 355 360		
GTC GAA TAC AGC GAA CAA AAA ACC ATC TTC TCT TGG TAGAGCAAGA		4015

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Val Glu Tyr Ser Glu Gln Lys Thr Ile Phe Ser Trp
365 370

GGCTGCCCTT GAAAGACTAA GAGCCACCCT GCCCTGTTTA AAGGGCTAAA AGTTTAATAT	4075
TTCTCTGCAG CCTAAACAGT TGGAAACATT GAAAATCTAG GATGTATCAG AAAAAAGAAG	4135
GTTTGGAGGA AGTATGGATG ATATAGAGGA CATGAATGTA TTCATTTTCG GTATACTCTT	4195
CTGCAAA ATAATTCTTC AGATGTTTTT GTGGTATGAG ATATAGAGGA CATGTATGTA	4255
TGCGGTAAGG CTGAAGTAAA CAAGTTACCA TAAGAGACAG CCCTCTCGGT TTCTTCCATC	4315
TGATCGATTC GTCTCGTCGA ATTTGCCAAA AGCTCAAAAC TCAACTCATC CCCTGCTTTC	4375
TATCCATATG GGCAAGGAAT ACAATTAGAC CAGTTTGATA CTTGTAATGA GAAGTTTAC	4434

(2) INFORMATION FOR ID SEQ NO:6

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2955 base pairs
- (B) TYPE: Nucleic acid
- (C) STRANDEDNESS: Double strand
- (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: No

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Cuphea lanceolata*

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: Genomic lambda FIX II
- (B) CLONE: ClGPDHg3

(ix) FEATURE:

- (A) NAME/KEY: CAAT signal
- (B) LOCATION: 1055 to 1058

(ix) FEATURE:

- (A) NAME/KEY: TATA signal
- (B) LOCATION: 1103 to 1107

(ix) FEATURE:

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(A) NAME/KEY: Start codon

(B) LOCATION: 1182 to 1184

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: Join (1182 to 1326, 1837 to 1913, 2010 to 2082, 2180 to 2397, 2480 to 2587, 2668 to 2731, 2848 to 2885, 2947 to 2955)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6

GGATCCTCCT CGATGGTGGT CCAATGAAGA CTATACAAA CCAAGCCGAC GGAATCCGGT	60
GCACAATAAC TTGAAGCCAT GAAAACCAAT GCAATATATA GAGTACGCCT TGTACTATGT	120
AATATATTTA CAATTTTCTC TTGAATAGTT TAGGTTTGGT GATCGTAAAC TCGCAAACA	180
CATATGTGCG TGTGTAA.ATA TATCTGGTGA TGATGTATGA AGAGAGTGCG GTTTAATTAC	240
CCGGTATTGT ATAAGGTTGT ATCTGCAGTT GACACTTTCA GTAGAAATTA CTAATAACTC	300
GACGAGATAC AAACGACTCG AGTTTCAGAA ATAAGTGGCA AAACGTTATG GGGTTCTCCT	360
TGATTCTTCG TGGAAGGTAT ACTATTAATC ATGTTTCGCCT CCGTCCTAGT AGAAACATAG	420
AGTTTTTATC GGGATGCAGA TTGCAGATGA TAGAACTATT GTCAGATTCA TTATGCATAT	480
AGGATAGGCC TTCTACTGAT TTGGAAACTT ATATCGATTG TGTGGAATG GATGTATGAA	540
AAGCTTCATA TCCGACATTG AAAATTTGGT CATATCAATA AGATGAACTA ACAAATATG	600
CCAACCTCTT GGAAGCAAAA CACATCCGAG ACTTTAAGAT GTGGCTGAGG TTTCTGCAAC	660
TTTAAATCTC CCATATGCTT GACAGAATTG GTAGACCTAA CTCAATGGAT TTCATTCAAT	720
GATCGAAGTT TCTCTATCGA TCATAGCTGT GAATTAGTAA GCAAATGTCC ATAATATATC	780
CCCGAAAACA CGTAAAGTTA GGTCTCATTA CATTAGGCCT CAACCATATG TTATAAGTAA	840
ATTTGTTTTT TTTTTTTTCT CTTACAGTTG AATGTATCAA ATCGAAAAA CCGTTAAGTC	900
GTTGCGGCCC TTTGAATAGT AAGCCAAAGA TCCGAAAGAA AAAGTAAACA GAGACAGAGC	960
AATGAGGAGA TGGCCAGTTT GAGAAGCAAA CGCATAGGTT GCCACGGAGG AGGCGGAGAC	1020
GGGTCATCGA TGACTTTCTC CGCCTCCTTA ACCGCAATGG CGATGCCGCC ATACCTCTCT	1080

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GTCACCCTCT CTCCATTCCC TTTATATCTC TCCCGCTTCT TCCTCTGCTC CACTCAACCC	1140
CCTCTGCATA AACTCTGTGC TTTTITAGTC TCTCCCCTGC T ATG TCG CCG GCA	1193
Met Ser Pro Ala	
1	
TTC GAA CCC CAT CAG CAG AAG CCT ACC ATG GAG AAC ATG CGA TTC CGA	1241
Phe Glu Pro His Gln Gln Lys Pro Thr Met Glu Asn Met Arg Phe Arg	
5 10 15 20	
GTC ACC ATC ATT G4GC AGC GGT AAC TGG GGC AGC GTC GCC GCT AAG CTC	1289
Val Thr Ile Ile Gly Ser Gly Asn Trp Gly Ser Val Ala Ala Lys Leu	
25 30 35	
ATT GCC TCC AAC ACC CTC AAC CTC CCG TCT TTC CAC G GTTTGTCTGC	1336
Ile Ala Ser Asn Thr Leu Asn Leu Pro Ser Phe His	
40 45	
CACTCTTCTT TCTTCATGAT CAGGCTCTTG CCAGTAGAGA CATGTCTTTT CATGAATCAA	1396
GCACCCGTTT TTTCGATGAG GATCACTGAG TTTGATTAA GGGTATCCGA TGCAACTGCT	1456
GAAAAGATGT GGTATTATTTT GTTCTTTCAT GAAGTATCAT CTGAGAAATT TGATCTTAGC	1516
CTAAGCGGCA TTAATTTTCGG TGTTAAGTTC ATTCTATGTG AGTAGGAGTA TGAGGTGATG	1576
CCGCGTGATT CCAATCAGGT ACCGATGAAA ATCAGTAGAC ATGGTTGCAG TTGAGGTTC	1636
ATAGTTTACA CAGCATAGGA GTTGCTGTAT TTCTATTGAC GCTTGGATTT GTTTGGTGCT	1696
TATAATCCCG GTTTTACTA ATTGGTTATG AACACCGATA ATAACAACAG TTAGATTTCT	1756
TCAACATTAA CCGGTTGAAG ATTAGGCCAT ATTCTTATTT GGGTACTATT TCTTAAGAAA	1816
ACATTTCATAT TTTCTTTCAG AT GAA GTA AGG ATG TGG GTG TTT GAG GAG	1865
Asp Glu Val Arg Met Trp Val Phe Glu Glu	
50 55	
ACA TTG CCA AGC GGC GAG AAG CTC ACT GAA GTC ATC AAC CGG ACC AAT	1913
Thr Leu Pro Ser Gly Glu Lys Leu Thr Glu Val Ile Asn Arg Thr Asn	
60 65 70	
GTAAGGAAGA TCAATTTAGC ATGTCATTGT ATTAACATAA AGAGCGTTTA TTGGCAACTT	1973
TGGCTTTCAT GATGTTTCGAG TGTTGCGTCT TTGCAG GAA AAT GTT AAG TAT CTG	2027
Glu Asn Val Lys Tyr Leu	
75 80	
CCT GGA TTC AAG CTT GGC AGA AAT GTT ATT GCA GAC CCA AAC CTT GAA	2075
Pro Gly Phe Lys Leu Gly Arg Asn Val Ile Ala Asp Pro Asn Leu Glu	
85 90 95	

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AAT GCA G GTAGTGATTG TATTTTCAGTG CTCGGTTGAA TGATCAAGTA AAATCCTCGT Asn Ala	2132
GCTAAATATG TCGAGATGTT CGTGTTTTTG CATAATGTTT TGTTTAG TT AAG GAA Val Lys Glu 100	2187
GCA AAC ATG CTT GTA TTT GTC ACA CCG CAT CAG TTC GTG GAG GGC CTT Ala Asn Met Leu Val Phe Val Thr Pro His Gln Phe Val Glu Gly Leu 105 110 115	2235
TGC AAG AGA CTC GTC GGG AAG ATA AAG GCA GGTGCA GAG GCT CTC TCC Cys Lys Arg Leu Val Gly Lys Ile Lys Ala Gly Ala Glu Ala Leu Ser 120 125 130	2283
CTT ATA AAG GGC ATG GAG GTC AAA AGG GAA GGG CCT TCC ATG ATA TCT Leu Ile Lys Gly Met Glu Val Lys Arg Glu Gly Pro Ser Met Ile Ser 135 140 145	2331
ACC TTA ATC TCG AGC CTT CTC GGG ATC AAC TGC TGT GTC CTA ATG GGA Thr Leu Ile Ser Ser Leu Leu Gly Ile Asn Cys Cys Val Leu Met Gly 150 155 160 165	2379
GCA AAC ATC GCC AAC GAG GTAAAATCTT GGTGCAGTCT TACGAGATTC Ala Asn Ile Ala Asn Glu 170	2427
TGAATCTTGA ACCTGTTAGC ATTTTGACAC ACTGTGACTT CTAAATTTGT AG ATT Ile	2482
GCT CTT GAG AAA TTC AGT GAG GCG ACA GTC GGA TAC AGA GAA AAT AAG Ala Leu Glu Lys Phe Ser Glu Ala Thr Val Gly Tyr Arg Glu Asn Lys 175 180 185	2530
GAT ACT GCA GAG AAA TGG GTT CGG CTC TTC AAC ACT CCA TAC TTC CAA Asp Thr Ala Glu Lys Trp Val Arg Leu Phe Asn Thr Pro Tyr Phe Gln 190 195 200	2578
GTC TCG TCT GTGAGTACGA ATAAACCTTT CCTTCTGCGA ACAAAAACT Val Ser Ser 205	2627
TCCCGAGGCA GGAAGTAAAT GAAACAAGTT AACATAATAG GTT CAA GAT GTG GAA Val Gln Asp Val Glu 210	2682
GGA GTG GAA CTT TGT GGC ACA CTG AAG AAT GTC GTG GCC ATA GCA GCC G Gly Val Glu Leu Cys Gly Thr Leu Lys Asn Val Val Ala Ile Ala Ala 215 220 225	2731
GTACTTATAT ACGATCTCCA CATTTATATA AACTAGTTAG AAAGATTTTG GATTGCTGTA	2791
AAAACCGTGG AAAAACCCGA AAAGTGTTGA TGAAGTGTTA CCAAATGTTG TTTCAG GT Gly	2849

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TTT GTA GAT GGA CTG GAG ATG GGA AAC AAC ACA AAG GTAAGTCCAA 2895
Phe Val Asp Gly Leu Glu Met Gly Asn Asn Thr Lys
230 235 240

AGTTCATGCA AATTTTTTCG TATTACGAC TGAATGCTTG GATATACATA G GCT GCG 2952
Ala Ala

ATT 2955
Ile

(2) INFORMATION FOR ID SEQ NO:7

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 574 base pairs
- (B) TYPE: Nucleic acid
- (C) STRANDEDNESS: Double strand
- (D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: No

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Cuphea lanceolata*

(vii) IMMEDIATE SOURCE:

(A) LIBRARY: Genomic lambda FIX II

(B) CLONE: ClGPDHg3

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 31 to 189

(ix) FEATURE:

(A) NAME/KEY: Stop codon

(B) LOCATION: 190 to 192

(ix) FEATURE:

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(A) NAME/KEY: PolyA signal

(B) LOCATION: 393 to 398

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7

GGCATATCGA TGATTTTCC TATCTTGCAG GGT GTC TTG ACA GCA AAA GAG GTG	54
Gly Val Leu Thr Ala Lys Glu Val	
1 5	
TAT GAG GTA CTG AAG CAC CGG GGC TGG CTC GAG CGT TTC CCG CTC TTC	102
Tyr Glu Val Leu Lys His Arg Gly Trp Leu Glu Arg Phe Pro Leu Phe	
10 15 20	
GCA ACT GTG CAT GAG ATC TCA TCT GGC AGG TTG CCT CCT TCA GCC ATT	150
Ala Thr Val His Glu Ile Ser Ser Gly Arg Leu Pro Pro Ser Ala Ile	
25 30 35 40	
GTC AAA TAC AGC GA-A CAA AAG CCC GTC TTA TCT CGA GGT TAGAACGAGA	199
Val Lys Tyr Ser Glu Gln Lys Pro Val Leu Ser Arg Gly	
45 50	
GAAAACCCGA CAAACCGGTG AAACCTCGTAG TCTTAAACTG AAATCCAAAA ACATGCTGGG	259
AACATCAGCA AAAACCATTC ATCAAGGATG TCTTAGATAA AAGGTTTCAG GAAGAAATAG	319
ATGGTAGTGT GTGTAATGTT ATCAGCAATC ATTCATTCAT TTATTAAGTA TTTTTCGCAT	379
CATATTTTAT GCTAATAATT ATTACATAAA TTAACCAAT TTTGTCAAAA TTTCTGCATT	439
GCCCCAAACA GATTAATGCA TTGAGAAAAA CTTATAAAGC TTTATCCAGC ATACATATAG	499
TTCTTTAAGC AATACAAAAA CACCCTTCTA AGCCTCTTTG AAGATGGAGT TTGATCACAC	559
ATTAAAATGC TTTT	574

(2) INFORMATION FOR ID SEQ NO:8

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1507 base pairs

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Double strand

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: DNA (molecular)

(iii) HYPOTHETICAL: No

(iv) ANTI-SENSE: No

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Cuphea lanceolata

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(vii) IMMEDIATE SOURCE:

(A) LIBRARY: Genomic lambda FIX II

(B) CLONE: ClGPDHg9

(ix) FEATURE:

(A) NAME/KEY: TATA signal

(B) LOCATION: 1108 to 1112

(ix) FEATURE:

(A) NAME/KEY: Start codon

(B) LOCATION: 1193 to 1193

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1193 to 1376

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8

GCATGCGGGC AGGCAGGCAG GCATGGGTCT AAATTCTAGA AGACCCAGAC ATATTCATTT	60
TGTTCAACAAC CGACCCATCA ATATATTGAT TAATTTTGTT TAAATTTATC ATCAGTTTTT	120
ATTTAATATT TTAAATAGG TTTACCTTGA TCGTGATAAT TATTTAATAT TACTTTGTAA	180
TAGTTTATTT ATCTAGCGTT ATAAAATAAC ATTTGAATTC GTTGATGATA TGTGTATTTT	240
TACTATGTTT ATATGAAATT TATATTTCAA ATATTAAATA ATGTTCTTAT TTTGGCCTAT	300
GGAGAAGTAT CATCAATTTT TCTATTAAAT AACAGTCTTC AGTTTAGTCA AATCAGTTGA	360
TAAGTTCCCA AATCACACAT TGTTTGTATG AAAATTTTAA TAAAAAAGTT AAGATGGTAT	420
TATTATAGAA AAATATATAA AGTATCTTTA AATAATAATT TCTTTTAAAT AAAAAAGGA	480
ATATTTGATT ACTTGACTTA TAAATTTTAT TGATAAGGAT GCCAACTTTC ATTTTAGAAA	540
CTAGAGTAAT GATGGTTAAA TTCCCCGAAA AATGGTATGT CAATTTATTG ATACGTTCCA	600
CTACTATTT CTGAGACATT TACATGTTTG TAAAAAAAAT CTATATATTT AAATTAAGAT	660
GGGTGTAATC AATTATAAAA TACAGCGAAT TTAAACACCG AATGAATAGA TTATCTGCAT	720
AACAATTTAT ACCATCCCTA AATACGAATT AGCAAGTTAA TAAATTTTAA TTACACGAAC	780

CATGATTATA TAAATTATCG AATCCCCGAC GTGGGGACGT ACCGAACCAA CCGTTGAAGT	840
GGTTGCCCTT TGAATCCTAA GACATACAGA CGTCATGATT CTTTGTCTCT CTATCTGTCC	900
ATTTACATAA TAAAATCAAA GAGAAGAAAA CAGAGGAAGC AGAGCATAGC ATAGCATAGC	960
ATAGAGGAGA TCGCCAGATT CAGCTGTTTC CTCATAGTTT GCCACGAGAC ATACATTGCA	1020
TTGCCCGATG CCTTTCTCCG CCTCCTTGTC CCTCTCCTCA TTCCCCGAT GCCTTTCTCC	1080
GCCTCCTTGT CCCTCTCCTC ATTCCCTTAT ATCCCTCCTC CCCTCCCTCT TCTTCCTCTG	1140
CTCAACTCCT CCCCCTCACC CTCTTCCTCT OTTCTTCCTC TCTGCCTCTG CA ATG	1195
	Met
	1
GCG CCT GCC TTC GAA CCC CAT CAG CTG GTT CCT TCT GAG CTT AAC TCT	1243
Ala Pro Ala Phe Glu Pro His Gln Leu Val Pro Ser Glu Leu Asn Ser	
5 10 15	
GCC CAC CAG AAC CCA CAT TCC AGC GGA TAT GAA GGA CCC AGA TCG AGG	1291
Ala His Gln Asn Pro His Ser Ser Gly Tyr Glu Gly Pro Arg Ser Arg	
20 25 30	
GTC ACC GTC GTT GGC AGC GGC AAC TGG GG4C AGC GTC GCT GCC AAG CTC	1339
Val Thr Val Val Gly Ser Gly Asn Trp Gly Ser Val Ala Ala Lys Leu	
35 40 45	
ATT GCT TCC AAC ACC CTC AAG CTC CCA TCT TTC CAT G GTTAGTCTCT	1386
Ile Ala Ser Asn Thr Leu Lys Leu Pro Ser Phe His	
50 55 60	
CATTCTTCTC TCTGTAAAGT TGAAGCTTTT TCATGGAATA GTCTCTAGAC ATGAGCCCCT	1446
GTTTGCATGG TTTTGTTTTG TCTTTGAAAC ATGAATAAAG GTGGTTTCTT GTGTTGGTAC	1506
c	1507

Patent Claims

1. DNA sequences which are isolated from plants and code for a glycerol-3-phosphate dehydrogenase, and the alleles as well as derivatives of these DNA sequences.
2. DNA sequences according to claim 1, wherein they are isolated from *Cuphea lanceolata*.
3. Genomic clones which are isolated from genomic plant DNA and contain a complete gene of a glycerol-3-phosphate dehydrogenase and the alleles as well as derivatives of this gene.
4. Genomic clones according to claim 3, wherein the complete gene contains the promoter sequence and other regulator elements in addition to the structure gene.
5. Genomic clones according to claim 4, wherein the plant DNA originated from *Cuphea lanceolata*.
6. Promoters and other regulator elements of the glycerol-3-phosphate gene from one of the genomic clones according to claims 3 to 5, and the alleles as well as the derivatives of these promoters.
7. DNA sequences according to claim 1, obtained from functional complementation with mutants of a microorganism.
8. DNA sequences according to claim 7, wherein the microorganism is *E. coli* BB26-36.
9. Procedure for producing plants, plant parts and plant products the triacylglyceride content or fatty acid pattern of which is altered, in connection with which a DNA sequence is transferred according to one of

claims 1 or 2, or a gene originating from the genomic clones according to one of claims 3 to 5 is transferred by genetic engineering methods.

10. Procedure according to claim 9, wherein the DNA sequence or the gene is transferred by microinjection, electroporation, particle gun, steeping of plant parts in DNA solutions, pollen or pollen tube transformation, transfer of corresponding recombinant Ti plasmids or Ri plasmids with *Agrobacterium tumefaciens*, liposome-mediated transfer, or by plant viruses.
11. Use of a DNA sequence according to one of claims 1 or 2 or of a gene originating from the genomic clones according to one of claims 3 to 5 for altering the biosynthesis output in plants.
12. Plants, plant parts and plant products produced according to a procedure of claims 9 or 10.

AMENDED PAGE

IPEA/EP